

How to Assess the Stage of Fibrosis in Chronic Hepatitis C

Detlef Schuppan

MECHANISMS OF PROGRESSION

Fibrosis and cirrhosis are a result of excess accumulation of extracellular matrix (ECM) molecules (collagens, noncollagenous glycoproteins, glycosaminoglycans, proteoglycans and of elastin [1]. Extensive perisinusoidal fibrosis has marked effects on liver function, due to the blockade of nutrient and metabolite exchange between hepatocytes and the circulation (Figure 1) and the liver is further impaired by the formation of novel intrahepatic vessels via porto-portal and porto-central collaterals that shunt the blood away from hepatocytes. The imbalance of two dynamic processes, fibrogenesis and fibrolysis leads to fibrosis. Activated hepatic stellate cells and myofibroblasts stimulate fibrogenesis by producing most ECM molecules, downregulating the expression of certain matrix metalloproteinases (MMPs), and increasing synthesis of physiological and tissue MMP inhibitors (TIMPs) [1-3] (Figure 2). Even advanced liver fibrosis and cirrhosis are reversible when the causes of fibrogenesis such as viral infection or biliary obstruction, are removed and the liver is given time to recover [4-11]. Furthermore a growing number of gene polymorphisms may either protect against or enhance the development of hepatic fibrosis (Table 1) [12-20]. In addition to the known external factors and the histological and serological markers of fibrosis and its development, these genetic polymorphisms

may provide individual risk profiles for the development of severe fibrosis.

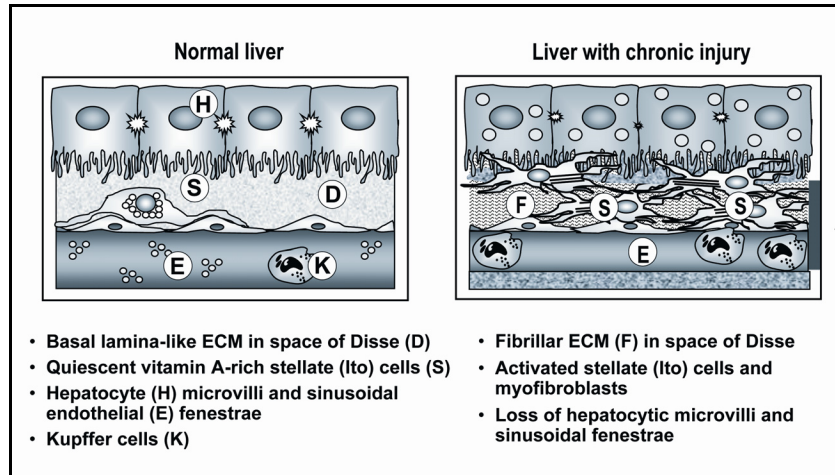


Figure 1: Capillarization of the sinusoids. Illustration of the major cell biological events that determine functionally relevant fibrosis [modified from a sketch kindly provided by Dr. M. Pinzani, Florence, Italy].

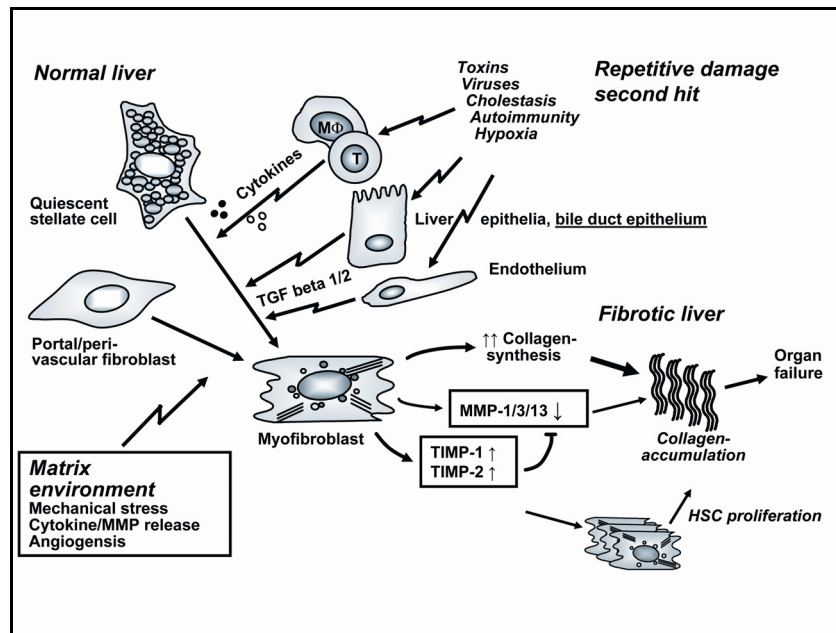


Figure 2: Initiation and maintenance of fibrogenesis. With continuous injury, primarily to hepatocytes or bile duct epithelia, and / or mechanical stress the normally quiescent hepatic stellate cells and portal/perivenular fibroblasts undergo activation and transdifferentiation to myofibroblasts. These myofibroblasts produce excessive amounts of collagens, downregulate certain MMPs and show an enhanced expression of the physiological inhibitors of the MMPs (TIMP-1 and -2). TIMP-1 can also promote myofibroblast proliferation and inhibit their apoptosis.

Gender (protection by high dose estrogens)
Pro/antioxidative enzyme polymorphisms (MnSOD, GSTP1, CYP2D6), e.g., in hemochromatosis
Immune system (profibrogenic Th2 vs. Th1 response)
Single nucleotide-polymorphisms (IL-1beta, IF-gamma, MCP-1, TNF-alpha, Factor V Leiden, MMP-3, TGF beta 1, DQB1*0503)
Genetically determined comorbidities: HFE mutations, metabolic syndrome (NASH)
Regulation of regeneration and apoptosis

Table 1: Genetic predisposition for hepatic fibrosis [12-20]. CYP2D6, cytochrome P450 2D6; GSTP1, glutathione S-transferase P1 [Stickel et al. unpublished data]; MnSOD, manganese superoxide dismutase [Oesterreicher et al. unpublished data]; NASH, non-alcoholic steatohepatitis.

IS THERE A GOLD STANDARD OF LIVER FIBROSIS?

Sequential histological grading of inflammation and particularly staging of fibrosis are still considered the gold standard to assess progression. However, certain studies have demonstrated sampling errors not only in patients with liver diseases with a high degree of intrahepatic heterogeneity such as biliary fibrosis, but also in patients with alcoholic or hepatitis C virus (HCV)-induced fibrosis and inflammation. Thus, when the well accepted, easy to use, 4 stage METAVIR score is used to stage fibrosis [21], roughly one third of the scores differed by at least one stage in the same patient when biopsies from the left and right liver lobes were compared [22]. Similar results were obtained when laparoscopic assessment of cirrhosis vs. non-cirrhosis (which is questionable as a gold standard) was matched to histological findings [23] (Table 2 and 3). Similar results were obtained for the grading of inflammation. This discrepancy was confirmed and systematically investigated in a recent study using the overall scoring of large surgical liver specimens from

patients with chronic hepatitis C as a gold standard. Results of this study showed that small, virtual biopsies derived from these large sections were correctly categorized in only 65% vs. 75% of cases when the biopsies were 15mm and 25mm long [24]. Moreover, a further increase in length from 25-45mm did not significantly increase accuracy. Therefore, although it is indispensable for many reasons, liver biopsy cannot be considered the ultimate gold standard for the assessment of stage and grade and thus the progression of fibrosis. This uncertainty complicates the search for non-invasive (serological) markers of the progression of fibrosis.

Homogeneity of staging & grading in chronic hepatitis C. HCV, laparoscopic biopsy of right and left liver n=124, METAVIR score		
Difference	n	%
≥1 stage	41/124	33.1
≥2 stages	3/124	2.4
≥1 grade	30/124	24.2
≥2 grade	2/124	1.6
cirrhosis vs. stage 3	18/124	14.5

Table 2: Sampling error in chronic hepatitis C [22].

Laparoscopy vs. Histology			
Retrospective, 1992-1994, 434 consecutive patients.			
HCV 52%, HBV 8%, FL 8%, PBC 4%, AIH 3%, others 25%			
	Laparoscopy	Histology	Error
Cirrhosis	169	115	32%
No cirrhosis	265	263	0.8%
Detection of cirrhosis (gold standard laparoscopy)			
Sensitivity of biopsy		68%	
Specificity of biopsy		0.8	

Table 3: Sampling error in chronic liver diseases [23].

IMAGING TECHNIQUES

At present imaging techniques lack the sensitivity and specificity necessary for the assessment of the stage of fibrosis in patients with chronic liver diseases. Structural, non-homogenous findings at ultrasound are not associated with the stage of fibrosis, and liver echogenicity can only be used for the detection or exclusion of moderate to extensive fatty infiltration [25]. Although the hepatic artery resistance index as measured by Doppler ultrasound was slightly higher in severe than in mild fibrosis, and no correlation was found with histological inflammation, necrosis or portal flow velocity, the method lacks sensitivity [26]. A slightly better differentiation between slight and severe fibrosis is found with magnetic resonance (MR)-techniques, such as superparamagnetic iron oxide-enhanced MR, which shows hypersignal intensities with a reticular pattern in most patients with advanced fibrosis (METAVIR F2-4), while the signal from non-fibrotic areas where more Kupffer cells are present is decreased [27]. The fibroscan, an interesting new technique using both ultrasound and low-frequency (50Hz) elastic waves whose propagation velocity are directly related to elasticity, was evaluated to

quantify liver fibrosis in 106 patients with chronic hepatitis C. The areas under the (ROC) curves were 0.88 and 0.99 for the diagnosis of significant fibrosis or cirrhosis (METAVIR F2-4 and F4) [28]. Further prospective studies are needed to determine whether this technique can be used to detect changes in the stage of fibrosis in individual patients, e.g. during antifibrotic therapy.

SEROLOGICAL MARKERS OF PROGRESSION

Several studies have been performed with combinations of known serum markers of synthetic, metabolic or excretory liver functions, to derive an algorithm that predicts the histological severity (stage and grade) of chronic liver diseases. These algorithms were retrospectively determined and prospectively validated. Examples are the fibroscore, using alpha 2-macroglobulin, haptoglobin, gamma glutamyl transferase (GGT), gamma-globulin and bilirubin [29-31], and another score using platelet count, GGT, age and cholesterol [32] in patients with chronic hepatitis C (Table 4 and 5). Although these scores can be used instead of liver biopsy in a certain number of patients when a decision to treat or not must be made, they do not appear to be suitable for scientific studies requiring greater accuracy and an assessment of the dynamics of fibrogenesis and fibrolysis. Thus, when making a treatment decision, simple indicators may suffice. For example a single increase in alanine aminotransferase (ALT) during a 6 month observation period in patients with chronic hepatitis C indicated \geq stage 1 fibrosis allowing treatment to begin. These results occurred in 90% of patients (Table 6) [33]. Other indices are the PGA (prothrombin time, GGT, apolipoprotein A with or without alpha 2-macroglobulin) which has been validated in patients with alcoholic liver disease (Table 7) [34, 35].

Non-connective tissue markers as predictors of relevant liver fibrosis in hepatitis C (Fibroscore)	
205 retrospective, 134 prospective patients with hepatitis C	
METAVIR F0-1 vs. F2-4	
5/11 serum markers predictive:	
	alpha-2 macroglobulin
	haptoglobin
	gamma-globulin
	GGT
	bilirubin
Index 0-0.1: 100% negative predictive of F2-4 (12%)	
Index 0.6-1.0: 90% positive predictive of F2-4 (34%)	
Index 0.1-0.6: no assignment possible (54%)	

Table 4: Diagnostic value of the fibroscore to predict fibrosis stage in patients with chronic hepatitis C [29].

Score to predict absent/little fibrosis (F0-1) in hepatitis C

351 retrospective, 125 prospective patients with hepatitis C
 METAVIR F0-1 vs. F2-4

Score: $7.811 - 3.131 \ln(\text{platelet count}) + 0.781 \ln(\text{GGT}) + 3.647 \ln(\text{age}) - 0.014(\text{cholesterol})$

Score <4.2	Stage 0-1	Stage 2-4
Estimation	120/266	5/125
Validation	47/92	2/49

Score >6.9	Stage 0-1	Stage 2-4
Estimation	10/47	37/85
Validation	5/15	10/33

Score <4.2: sensitivity 51%, NPV 96%

Score >6.9: sensitivity 30%, PPV 66%

Table 5: Alternative index for prediction of fibrosis in patients with chronic hepatitis C [32].

Prediction of absent/little fibrosis (F0-1) by ALT					
864 retrospective patients with hepatitis C					
METAVIR F0-1 vs. F2-4					
ALT normal vs. ALT elevated during 6 months					
Stage	0	1	2	3	4
ALT normal	34.8	51.5	12.1	0	1.5
ALT elevated	0.8	23.7	50.5	17	8
ALT persistently normal (n=66): 65% ≥F1, 26% >A1F1					
ALT elevated: 99% ≥F1, 88% >A1F1					
Cut-off ALT >2.25 ULN: clear indication for treatment					
All patients with elevated ALT can be treated					
Biopsy only for patients with normal ALT					
Table 6: ALT as a predictor of relevant fibrosis or inflammation in patients with chronic hepatitis C [33].					

PGA- or PGAA-index and alcoholic liver disease

Patients with alcoholic liver disease:
n=333 retrospective, n=291 prospective

METAVIR F0-1 vs. F4

Serum markers: prothrombin time

gamma GT

apolipoprotein A

Index 0-2: 100% neg. pred. for F3/F4, 83% pos. pred. for F0/F1

Index 9-12: 0% neg. pred. for F0/F1, 86% pos. pred. for F3/F4

Correct classification of 65% of patients (Poynard et al. 1991 [34])

PGAA-Index (incl. alpha-2 macroglobulin n=316 prospective):

Correct classification of 70% of patients (Naveau et al. 1994 [35])

Table 7: PGAA and PGA indices to predict the severity of alcoholic liver fibrosis [34-35].

Measuring circulating metabolites of the ECM appears to be a more straightforward approach to assess fibrogenesis and fibrolysis, especially in studies on the inhibition or reversal of liver fibrosis (Figure 3) [36-38]. However, serum levels of these markers are influenced by their excretion via the kidney or in bile, and by their uptake by endothelial cells, especially by liver sinusoidal endothelial cells. In addition, other organs with a high ECM turnover can contribute to these serum levels. Cross-sectional studies suggest a significant, but insufficient predictive value of single ECM markers for the stage of fibrosis [39-41]. Meanwhile the cross-sectional evaluation of the European liver fibrosis consortium (ELF) study using 10 automatized ECM parameters in more than 1000 patients with various chronic liver diseases provided algorithms of 3-4 ECM markers with a better predictive value than an assessment by an independent expert pathologist who was not trained as well as two reference pathologists [42]. As in other studies correlating histology

with noninvasive markers, the problem of validation for bioptical sampling errors remains (see chapter above) which introduces an error of one stage (METAVIR scale) in 25% of biopsies; this is expected to increase when liver diseases other than chronic hepatitis C are included (as in the ELF study). The results of the two-year follow-up arm of the ELF study have not yet been published.

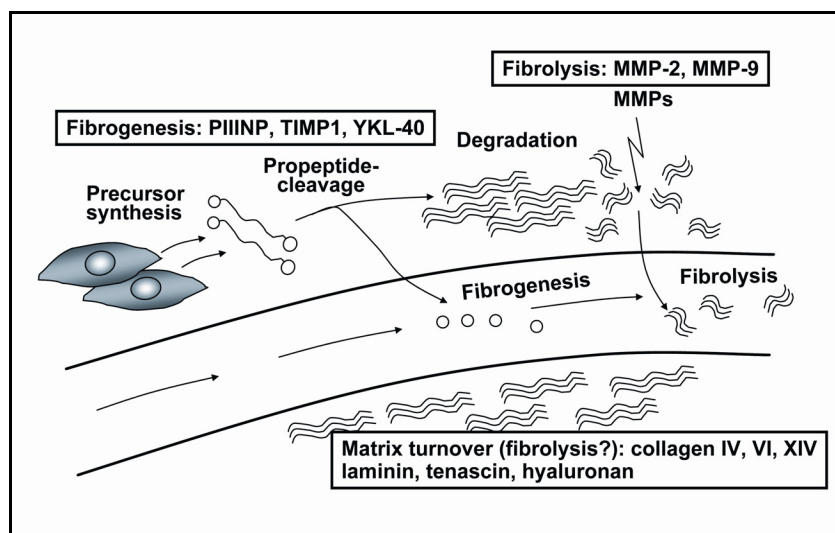


Figure 3: Circulating matrix proteins related to fibrogenesis and fibrolysis. Procollagen precursors released by fibrogenic cells are processed by procollagen peptidases. Only removal of the bulky propeptides allows the formation of collagen fibrils in the extracellular space. Thus circulating propeptide levels should reflect de novo synthesis and deposition of collagen, i.e. fibrogenesis. On the other hand, action of MMPs is expected to generate fragments of already deposited matrix proteins the levels of which should reflect matrix dissolution, i.e. fibrolysis. Most other molecules appear to rather represent an accelerated matrix turnover. The two large multicenter studies that evaluate the predictive value of circulating matrix markers as predictors of fibrosis stage are mentioned (ELF: patients with all chronic liver diseases; Prometheus: patients with chronic hepatitis C). The ELF study also assesses the predictive value as to fibrosis progression.

A more direct approach to validate the true serum markers of fibrogenesis and fibrolysis, which is nevertheless equally prone to sampling errors, is the use of real time quantification of fibrosis-relevant mRNA expression from liver biopsies compared to serum fibrosis markers. In a study of 50 patients with various types of liver disease, we found a fairly good correlation between liver procollagen I or TIMP-1 expression and serum levels of the aminoterminal procollagen type III peptide or TIMP-1 (data not shown). These results need to be confirmed in larger studies. The availability of serum markers of hepatic fibrogenesis (or fibrolysis) will provide a quick and frequent assessment of the antifibrotic potential of drugs in patients with progressive liver disease. If these reliable serological tests can be combined with drugs that inhibit or revert fibrosis [43] the desire to revert fibrosis or even cirrhosis may be fulfilled.

REFERENCES

1. Schuppan D, Ruehl M, Somasundaram R, Hahn EG. Matrix as a modulator of hepatic fibrogenesis. *Sem Liver Dis* 2001;21:351-372.
2. Friedman SL. Molecular regulation of hepatic fibrosis, an integrated cellular response to tissue injury. *J Biol Chem* 2000;275:2247-2250.
3. Benyon RC, Arthur MJ. Extracellular matrix degradation and the role of hepatic stellate cells. *Semin Liver Dis* 2001;21:373-384.
4. Soyer MT, Ceballos R, Aldrete JS. Reversibility of severe hepatic damage caused by jejunoileal bypass after re-establishment of normal intestinal continuity. *Surgery* 1976;79:601-604.
5. Dufour JF, DeLellis R, Kaplan MM. Reversibility of hepatic fibrosis in autoimmune hepatitis. *Ann Intern Med* 1997;127:981-985.
6. Hammel P, Couvelard A, O'Toole D, Ratouis A, Sauvanet A, Flejou JF, Degott C, Belghiti J, Bernades P, Valla D, Ruzsniwski P, Levy P. Regression of liver fibrosis after biliary drainage in patients with chronic pancreatitis and stenosis of the common bile duct. *N Engl J Med* 2001;344:418-423.
7. Muretto P, Angelucci E, Lucarelli G. Reversibility of cirrhosis in patients cured of thalassemia by bone marrow transplantation. *Ann Intern Med* 2002;136:667-672.
8. Crone J, Moslinger D, Bodamer OA, Schima W, Huber WD, Holme E, Stockler Ipsiroglu S. Reversibility of cirrhotic regenerative liver nodules upon NTBC treatment in a child with tyrosinaemia type I. *Acta Paediatr* 2003;92:625-628.
9. Poynard T, McHutchison J, Manns M, Trepo C, Lindsay K, Goodman Z, Ling MH, Albrecht J. Impact of pegylated interferon alfa-2b and ribavirin on liver fibrosis in patients with chronic hepatitis C. *Gastroenterology* 2002;122:1303-1313.
10. Arif A, Levine RA, Sanderson SO, Bank L, Velu RP, Shah A, Mahl TC, Gregory DH. Regression of fibrosis in chronic hepatitis C after therapy with interferon and ribavirin. *Dig Dis Sci* 2003;48:1425-1430.
11. Arthur MJ. Reversibility of liver fibrosis and cirrhosis following treatment for hepatitis C. *Gastroenterology* 2002;122:1525-1528.
12. Powell EE, Edwards-Smith CJ, Hay JL, Clouston AD, Crawford DH, Shorthouse C, Purdie DM, Jonsson JR. Host genetic factors influence disease progression in chronic hepatitis C. *Hepatology* 2000;31:828-833.
13. Bahr MJ, el Menuawy M, Boeker KH, Musholt PB, Manns MP, Lichtinghagen R. Cytokine gene polymorphisms and the susceptibility to liver cirrhosis in patients with chronic hepatitis C. *Liver Int* 2003;23:420-425.

14. Muhlbauer M, Bosserhoff AK, Hartmann A, Thasler WE, Weiss TS, Herfarth H, Lock G, Scholmerich J, Hellerbrand C. A novel MCP-1 gene polymorphism is associated with hepatic MCP-1 expression and severity of HCV-related liver disease. *Gastroenterology* 2003;125:1085-1093.
15. Hellier S, Frodsham AJ, Hennig BJ, Klenerman P, Knapp S, Ramaley P, Satsangi J, Wright M, Zhang L, Thomas HC, Thursz M, Hill AV. Association of genetic variants of the chemokine receptor CCR5 and its ligands, RANTES and MCP-2, with outcome of HCV infection. *Hepatology* 2003;38:1468-1476.
16. Satsangi J, Chapman RW, Haldar N, Donaldson P, Mitchell S, Simmons J, Norris S, Marshall SE, Bell JI, Jewell DP, Welsh KI. A functional polymorphism of the stromelysin gene (MMP-3) influences susceptibility to primary sclerosing cholangitis. *Gastroenterology* 2001;121:124-130.
17. Wright M, Goldin R, Hellier S, Knapp S, Frodsham A, Hennig B, Hill A, Apple R, Cheng S, Thomas H, Thursz M. Factor V Leiden polymorphism and the rate of fibrosis development in chronic hepatitis C virus infection. *Gut* 2003;52:1206-1210.
18. Yoshizawa K, Ota M, Saito S, Maruyama A, Yamaura T, Rokuhara A, Orii K, Ichijo T, Matsumoto A, Tanaka E, Kiyosawa K. Long-term follow-up of hepatitis C virus infection: HLA class II loci influences the natural history of the disease. *Tissue Antigens* 2003;61:159-165.
19. Erhardt A, Maschner-Olberg A, Mellenthin C, Kappert G, Adams O, Donner A, Willers R, Niederau C, Haussinger D. HFE mutations and chronic hepatitis C: H63D and C282Y heterozygosity are independent risk factors for liver fibrosis and cirrhosis. *J Hepatol* 2003;38:335-342.
20. Silvestri L, Sonzogni L, De Silvestri A, Gritti C, Foti L, Zavaglia C, Leverì M, Cividini A, Mondelli MU, Civardi E, Silini EM. CYP enzyme polymorphisms and susceptibility to HCV-related chronic liver disease and liver cancer. *Int J Cancer* 2003;104:310-317.
21. Bedossa P, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. The METAVIR Cooperative Study Group. *Hepatology* 1996;24:289-293.
22. Regev A, Berho M, Jeffers LJ, Milikowski C, Molina EG, Pyrsopoulos NT, Feng ZZ, Reddy KR, Schiff ER. Sampling error and intraobserver variation in liver biopsy in patients with chronic HCV infection. *Am J Gastroenterol* 2002;97:2614-2618.
23. Poniachik J, Bernstein DE, Reddy KR, Jeffers LJ, Coelho-Little ME, Civantos F, Schiff ER. The role of laparoscopy in the diagnosis of cirrhosis. *Gastrointest Endosc* 1996;43:568-571.
24. Bedossa P, Dargere D, Paradis V. Sampling variability of liver fibrosis in chronic hepatitis C. *Hepatology* 2003;38:1449-1457.
25. Mathiesen UL, Franzen LE, Aselius H, Resjo M, Jacobsson L, Foberg U, Fryden A, Bodemar G. Increased liver echogenicity at ultrasound examination

- reflects degree of steatosis but not of fibrosis in asymptomatic patients with mild/moderate abnormalities of liver transaminases. *Dig Liver Dis* 2002;34:516-522.
26. Sandrin L, Fourquet B, Hasquenoph JM, Yon S, Fournier C, Mal F, Christidis C, Ziol M, Poulet B, Kazemi F, Beaugrand M, Palau R. Transient elastography: a new noninvasive method for assessment of hepatic fibrosis. *Ultrasound Med Biol* 2003;29:1705-1713.
 27. Lucidarme O, Baleston F, Cadi M, Bellin MF, Charlotte F, Ratziu V, Grenier PA. Non-invasive detection of liver fibrosis: Is superparamagnetic iron oxide particle-enhanced MR imaging a contributive technique? *Eur Radiol* 2003;13:467-474.
 28. Sandrin L, Fourquet B, Hasquenoph JM, Yon S, Fournier C, Mal F, Christidis C, Ziol M, Poulet B, Kazemi F, Beaugrand M, Palau R. Transient elastography: a new noninvasive method for assessment of hepatic fibrosis. *Ultrasound Med Biol* 2003;29:1705-1713.
 29. Imbert-Bismut F, Ratziu V, Pieroni L, Charlotte F, Benhamou Y, Poynard T; MULTIVIRC Group. Biochemical markers of liver fibrosis in patients with hepatitis C virus infection: a prospective study. *Lancet* 2001;357:1069-1075.
 30. Poynard T, McHutchison J, Manns M, Myers RP, Albrecht J. Biochemical surrogate markers of liver fibrosis and activity in a randomized trial of peginterferon alfa-2b and ribavirin. *Hepatology* 2003;38:481-492.
 31. Myers RP, Benhamou Y, Imbert-Bismut F, Thibault V, Bochet M, Charlotte F, Ratziu V, Bricaire F, Katlama C, Poynard T. Serum biochemical markers accurately predict liver fibrosis in HIV and hepatitis C virus co-infected patients. *AIDS* 2003;17:721-725.
 32. Forns X, Ampurdanes S, Llovet JM, Aponte J, Quinto L, Martinez-Bauer E, Bruguera M, Sanchez-Tapias JM, Rodes J. Identification of chronic hepatitis C patients without hepatic fibrosis by a simple predictive model. *Hepatology* 2002;36:986-992.
 33. Pradat P, Alberti A, Poynard T, Esteban JI, Weiland O, Marcellin P, Badalamenti S, Trepo C. Predictive value of ALT levels for histologic findings in chronic hepatitis C: a European collaborative study. *Hepatology* 2002;36:973-977.
 34. Poynard T, Aubert A, Bedossa P, Abella A, Naveau S, Paraf F, Chaput JC. A simple biological index for detection of alcoholic liver disease in drinkers. *Gastroenterology* 1991;100:1397-1402.
 35. Naveau S, Poynard T, Benattar C, Bedossa P, Chaput JC. Alpha-2-macroglobulin and hepatic fibrosis. Diagnostic interest. *Dig Dis Sci* 1994;39:2426-2432.
 36. Schuppan D, Stölzel U, Oesterling C, Somasundaram R. Serum markers for liver fibrosis. *J Hepatol* 1995;22:82-88.

37. Oh S, Afdhal NH. Hepatic fibrosis: are any of the serum markers useful? *Curr Gastroenterol Rep* 2001;3:12-18.
38. Trinchet JC. Clinical use of serum markers of fibrosis in chronic hepatitis. *J Hepatol* 1995;22:89-95.
39. Nojgaard C, Johansen JS, Krarup HB, Holten-Andersen M, Moller A, Bendtsen F; Danish Viral Hepatitis Study Group. Effect of antiviral therapy on markers of fibrogenesis in patients with chronic hepatitis C. *Scand J Gastroenterol* 2003;38:659-665.
40. Stickel F, Poeschl G, Schuppan D, Conradt C, Strenge-Hesse A, Fuchs FS, Hofmann WJ, Seitz HK. Serum hyaluronate correlates with histological progression in alcoholic liver disease. *Eur J Gastroenterol Hepatol* 2003;15:945-950.
41. Patel K, Lajoie A, Heaton S, Pianko S, Behling CA, Bylund D, Pockros PJ, Blatt LM, Conrad A, McHutchinson JG. Clinical use of hyaluronic acid as a predictor of fibrosis change in hepatitis C. *J Gastroenterol Hepatol* 2003;18:253-7.
42. Rosenberg WMC, Voelker M, Thiel R, Becka M, Burt, Schuppan D, Hubscher S, Roskams T, Pinzani M, Arthur MJP, and the European Liver Fibrosis Group A cohort study of serum markers as predictors of human hepatic fibrosis assessed with histology and image analysis, submitted.
43. Schuppan D, Strobel D, Hahn EG. Hepatic fibrosis - therapeutic strategies. *Digestion* 1998;59:385-390.

